

Plant Extracts and Terpenes with Antivenom Properties

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Abstract

Envenoming and snakebite mortality are a significant public health concern in rural tropical areas in developing countries due to limited access to healthcare. Anti-snake venom (ASV) is the most effective counter-measure for treating snake bites. The antivenom is unavailable to the rural folk due to its high cost and poor health systems. Medicinal plants are a rich source of bioactive compounds with therapeutic activities and have been reported to exhibit antivenom properties. The aim of this study was to provide a review of antivenom plant extracts and terpenes. From the accessed literature information, 345 plants were reported to exhibit antivenom properties against snake venoms. However, the information on identification of the antivenom principles is scanty. Most of the plants investigated belong to the Fabaceae, Euphorbiaceae, Apocynaceae, Asteraceae, Lamiaceae and Rubiaceae plant families. Terpenoids reported to have antivenom properties include β -sitosterol, stigmasterol, lupeol, lupeol acetate, lupenone, 28-hydroxylupenone, betulin, betulinic acid, friedelin, β -amyrin, arjunolic acid, oleanolic acid, 11-deoxoglycyrrhetic acid, quinovic acid and ursolic acid. The results from this study show that plant extracts possess potent snake venom neutralizing activity and could provide an alternative way to inhibit venom toxins in snakebite. Further research to isolate and characterize the antivenom principles from the medicinal plants, and make antivenom formulations is necessary.

Keywords: Snake venom; antivenom; plant extract; terpenes

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I. Introduction

Envenomation from snakebite is a severe and often ignored public health issue that affects people all around the world, and particularly in impoverished nations¹⁻³. High concentrations of toxic proteins found in snake venom cause inflammation, bleeding, necrosis, neurotoxicity, cytotoxicity, cardiotoxicity, and ultimately mortality⁴. Every year, between 1.2 and 5.5 million snakebite incidents occur worldwide resulting in 25,000 to 125,000 fatalities⁵. Due to limited access to healthcare caused by poverty, envenoming and snakebite mortality are a significant public health concern in rural tropical areas in developing countries⁶.

Currently, the conventional anti-snake venom (ASV) is the most effective counter-measure for treating snake bites. However, the antivenom is hardly available in rural areas due to its expensive price and unfavorable storage conditions requirement⁷. Additionally, not many hospitals or institutions dedicated to managing snake bites are available in rural areas. Therefore, it is necessary to search for a novel, affordable and readily available remedy for snake envenomation^{1,6}.

Medicinal plants play an important role in healthcare since they are a rich source of bioactive compounds with therapeutic activities⁸⁻¹⁰. A large proportion of population in developing countries still depend on herbal medicine for their basic medical care¹¹⁻¹⁵. Plant-produced secondary metabolites have anti-disease characteristics for a variety of illnesses¹⁶⁻²⁰. In most cases, traditional remedies are favored because of their accessibility, effectiveness, affordability and lack of adverse effects to the user and environment^{11, 21-25}. Furthermore, there is little likelihood of acquiring medication resistance¹¹. Currently, different medicinal plants are being investigated through *in-vivo* and *in-vitro* trials to ascertain claims on their therapeutic properties²⁶⁻³⁰. Previous phytochemical analysis of bioactive extracts from plants has led to discovery of a number of important bioactive drugs including terpenoids, alkaloids, steroids, flavonoids and quinones³¹⁻³⁶. A number of secondary metabolites from plants that had not previously been known to have pharmacological activities are also being studied as a source of medicinal agents³⁷⁻⁴¹. Such bioactive compounds are a major source of lead compounds that are useful in new drug discovery and drug development⁴²⁻⁵¹. This study provides a review on plant extracts and terpenes with snake antivenom properties.

II. Antivenom Plant Extracts

Extracts from various plants have been tested for therapeutic activity againsts snake venoms. From the data accessed/used in this study, 345 plant extracts were reported to have antivenom properties against various snake venoms. The most investigated plants belong to the following plant families (the numbers in brackets

indicate the number of plants investigated): Fabaceae (41), Euphorbiaceae (25), Apocynaceae (18), Asteraceae (16), Lamiaceae (14), Rubiaceae (11), Zingiberaceae (9), Anacardiaceae (8), Annonaceae (8), Bignoniaceae (8), Moraceae (8), and Solanaceae (8) (Figure 1). These studies revealed that plant extracts inhibit enzymes in snake venom and act as antidotes^{2, 52, 53}. For example, extracts of *Indigofera pulchra* and *Aristolochia albida* gave 33.3% and 44.4% protection to mice treated with minimum lethal dose of venom by neutralizing the anticoagulant, hemolytic, and phospholipase activity of crude venom⁵⁴. *Curcuma aromatica*, *Aristolochia indica*, *Androgrphis paniculata* and *Curcuma zeodaria* extracts significantly antagonized *Daboia russelli*, *Echis carinatus*, *Ophiophagus hannah* and *Naja kaouthia* venom-induced lethal activity both in *in vitro* and *in vivo* studies⁵⁵. In another study, hydroethanolic root extract of *Solanum xanthocarpum* significantly neutralized the venom enzymes of *Naja*⁵⁶. A summary of some plant extracts that have been investigated, plant families, part of plant used and snake venom is provided in Table 1. Phytochemical analysis of the bioactive compounds in extracts of the most cited antsnake bite plants revealed the presence of alkaloids, flavonoids, tannins, aponosides and terpenoids⁵⁷.

According to previous reports, snakes with major clinical importance are those from the families Elapidae (African and Asian cobras, Asian kraits, African mambas, American coral snakes, Australian and New Guinean venomous snakes, and sea snakes) and Viperidae (Old World vipers, American rattlesnakes and pit vipers, and Asian pit vipers)⁵⁸. From the data accessed in this study the most tested venoms were obtained from snakes belonging to the genus *Naja* (frequency = 240) and *Bothrops* (frequency = 145). *Snake venoms from Bitis, Echis, Daboia, Lachesis, Crotalus* and *Bungarus* genus have also been investigated (Figures 2&3).

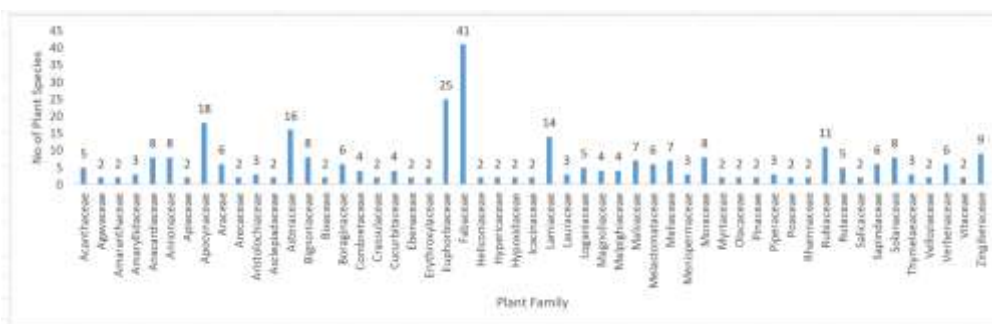


Figure 1. Some plant families studied for snake antivenom activity

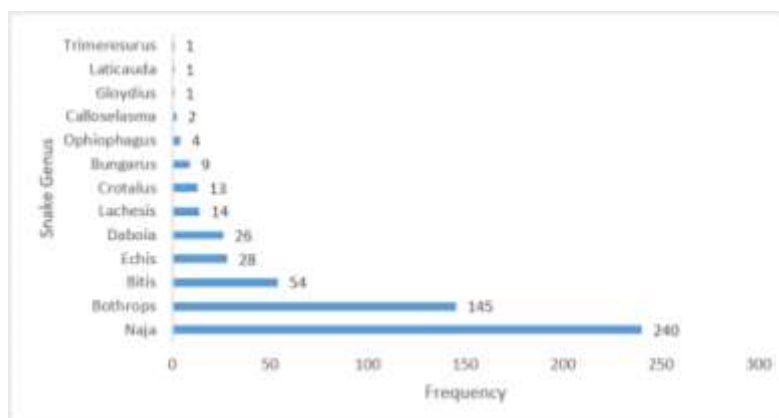


Figure 2: Some most commonly studies snake genus

Araceae	<i>Acorus calamus</i>	Roots		98
	<i>Colocasia esculenta</i>	Tuber	<i>N. nigricollis</i>	71
	<i>Dracontium croatii</i>	Rhizome	<i>B. asper</i>	99
	<i>Penellia ternate</i>	Rhizome	<i>B. jaracaca</i>	100
	<i>Philodendron megalophyllum</i>	Liana, vine	<i>B. atrox, B. jararaca</i>	67, 101
	<i>Phylodendrum tripatitum</i>	Branch, leaf	<i>V. russelli, B. atrox</i>	102, 103
Araliaceae	<i>Polyscias fulva</i>	Cortex	<i>N. nigricollis, B. arietans</i>	71
Arecaceae	<i>Areca catechu</i>	Seeds	<i>Bungarus caeruleus</i>	52
	<i>Borassus aethiopum</i>	Fruit	<i>N. mossandica</i>	104
Aristolochiaceae	<i>Aristolochia indica</i>	Root, Leaves, Whole plant	<i>D. russellii, E. carinatus, N. kaouthia, O. hannah, B. atrox, N. naja</i>	55, 59, 105
	<i>Aristolochia odoratissima</i>	Leaves	<i>B. atrox, N. naja and V. russelli</i>	106, 105
	<i>Aristolochia bracteolata</i>	Leaf, root	<i>N. naja, D. russelli</i>	97
Asclepiadaceae	<i>Hemidesmus indicus</i>	Roots	<i>V. russelli, B. atrox, E. carinatus, N. naja, Bungarus caeruleus, D. russelli</i>	55, 59, 102, 103, 107, 108
Asclepiadaceae	<i>Pergularia daemia</i>	Stem Bark	<i>N. naja</i>	109
Asteraceae	<i>Artemisia absinthium</i>	Aerial parts	<i>Montivipera xanthina</i>	110
	<i>Artemisia campstresis</i>		<i>Macrovipera lebetina</i>	111
	<i>Bidens pilosa</i>	Leaves, whole part	<i>Dendroaspis jamesoni and E. ocellatus</i>	112
	<i>Callilepis laureola</i>	Radix	<i>N. nigricollis</i>	71
	<i>Chaptalia nutans</i>	Leaf	<i>B. asper</i>	113
	<i>Cynara scolymus</i>	Leaves	<i>B. jaracaca</i>	114
	<i>Echinacea purpurea</i>	Roots	<i>B. asper</i>	115
	<i>Eclipta prostrata</i>	Roots, aerial parts, herbs	<i>C. durissus, C. d. terrificus, Calloselasma rhodostoma, B. jararaca, B. jararacussu, L. muta, Malayan pit viper</i>	105, 107, 116
	<i>Enhydra fluctuans</i>	Leaf	ND	69
	<i>Mikania glomerata</i>	Leaves, stem, roots	<i>B. jararaca, L. muta, Crotalus spp</i>	89, 114, 117, 118
	<i>Mikania laevigata</i>	Leaf	<i>Philodryas olfersii</i>	119
	<i>Neurolaena lobata</i>	Leaf, stem	<i>B. atrox</i>	81
	<i>Pluchea indica</i>	Root	<i>E. carinatus, N. naja, Bungarus caeruleus, D. russellii, N. kaouthia</i>	55, 120
	<i>Pseudelephantopus spicatus</i>	Whole plant	<i>B. atrox</i>	81
	<i>Elephantopus scaber</i>	Leaves	<i>B. jaracaca</i>	114
	<i>Vernonia condensata</i>	Leaves	<i>B. jaracaca</i>	114
	Balanitaceae	<i>Balanites aegyptiaca</i>	Stem bark	<i>E. carinatus</i>
<i>Basella alba</i>		Fruit	<i>B. atrox</i>	106, 112
Bignoniaceae	<i>Fridericia chica</i>	Leaf	<i>B. atrox, C. d. ruruima</i>	122
	<i>Jacaranda decurrens</i>	Leaves, roots,	<i>B. jararacussu, C. d. terrificus</i>	123
	<i>Kigelia africana</i>	Cortex, folium	<i>N. nigricollis, B. arietans</i>	71
	<i>Newbouldia leavis</i>	Leaves	<i>N. melanoleuca</i>	65
	<i>Radermachera xylocarpa</i>	Fruit	ND	69
	<i>Stenolobium stans</i>	Roots	<i>N. naja karachiensis</i>	72
	<i>Tabebuia aurea</i>	Stem bark	<i>B. newwiedi, V. russelli, B. atrox, B. asper</i>	81, 99, 103
	<i>Tecoma stans</i>		<i>N. n. karachiensis</i>	72
Bixaceae	<i>Bixa orellana</i>	Branch, leaf	<i>V. russelli, B. atrox</i>	102, 99
	<i>Cochlospermum tinctorium</i>	Radix	<i>B. arietans</i>	71
Boraginaceae	<i>Argusia argentea</i>	Bark	<i>T. flavoviridis, G. blomhoffii, B. arietans</i>	124
	<i>Cordia macleodii</i>	Bark	<i>N. naja</i>	125, 126
	<i>Cordia nodosa</i>	Bark	<i>N. atra, B. asper, B. atro</i>	127
	<i>Cordia verbenacae</i>	Leaves	<i>B. jararacussu</i>	128
	<i>Ehretia buxifolia</i>	Rootbark	<i>E. carinatus</i>	129
	<i>Trichodesma indicum</i>	Whole plant	<i>N. naja karachiensis</i>	72
Burseraceae	<i>Boswellia delzielli</i>	Stem Bark		130
Capparaceae	<i>Capparis tomentosa</i>	Radix	<i>N. nigricollis, B. arietans</i>	71
Capparidaceae	<i>Crateva magna</i>	Twigs, flowers	<i>N. nigricollis</i>	60
Caricaceae	<i>Carica papaya</i>	Leaves	ND	131

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Celastraceae	<i>Maytenus senegalensis</i>	Root	<i>N. naja</i>	132
Cleomaceae	<i>Cleome gynandra</i>	Leaves	<i>N.naja</i>	133, 134
Clusiaceae	<i>Clusia fluminensis</i>	Leaves, stems, flowers, fruits	<i>B. jararaca, B. jararaca and B. jararacussu</i>	135, 136
Colchicaceae	<i>Gloriosa superba</i>	Twigs, flowers, radix	<i>N. nigricollis, B. arietans</i>	71, 60
Combretaceae	<i>Combretum leprosum</i>	Root	<i>B. jararaca, B. jararacussu</i>	137
	<i>Combretum molle</i>	Folium	<i>N. nigricollis, B. arietans</i>	71
	<i>Guiera senegalensis</i>	Radix	<i>N. nigricollis, B. arietans, E. carinatus</i>	71, 138
	<i>Terminalia arjuna</i>	Bark	<i>N. naja karachiensis</i>	72
Connaraceae	<i>Connarus favosus</i>	Bark	<i>B. atrox, B. jararaca</i>	67, 101
Convolvulaceae	<i>Ipomoea rubens</i>	Seed	<i>N. nigricollis</i>	71
Crassulaceae	<i>Bryophyllum pinnatum</i>	Leaf	<i>B. jararaca</i>	139
	<i>Kalanchoe laciniata</i>	Leaf	<i>B. jararaca</i>	67
Cucurbitaceae	<i>Citrullus colocynthis</i>	Whole plant, fruit	<i>N. naja karachiensis, N. nigricollis</i>	72, 140
	<i>Luffa cylindrica</i>	Leaf	<i>N. naja</i>	141
	<i>Momordica charantia</i>	Fruits	<i>N. naja karachiensis</i>	72
	<i>Wilbrandia ebracteata</i>	Roots	<i>B. jaracaca</i>	114
Cyperaceae	<i>Scleria pterota</i>	Leaves	<i>B. jararacussu, B.moojeni, B.nalernatus,</i>	142
Dicranaceae	<i>Dicranum frigidum</i>	Whole plant	<i>B. asper</i>	74
Dilleniaceae	<i>Davilla elliptica</i>	Leaf	<i>B. jararaca</i>	143
Dryopteridaceae	<i>Dryopteris cochleata</i>	rhizome	ND	69
Ebenaceae	<i>Diospyros kaki</i>	Fruits	<i>Laticauda semifasciata</i>	144, 145
	<i>Diospyros mespiliformis</i>	Cortex	<i>N. nigricollis, B. arietans</i>	71
Erythroxylaceae	<i>Erythroxylum ovalifolium</i>	Stem	<i>L. muta</i>	146, 147
	<i>Erythroxylum subsessile</i>	Stem	<i>L. muta</i>	146, 147
Euphorbiaceae	<i>Abrus precatorius</i>	Radix	<i>N. nigricollis</i>	71
	<i>Acalypha Fruticosa</i>	Leaves	<i>E. species</i>	148
	<i>Acalypha indica</i>	Leaves	<i>Viper russelli</i>	149
	<i>Alchornea laxiflora</i>	Cortex	<i>N. nigricollis, B. arietans</i>	71
	<i>Argyrobolium stipulaceum</i>	Radix	<i>N. nigricollis</i>	71
	<i>Bridelia ferruginea</i>	Leaves	<i>N. nigricollis</i>	150
	<i>Burkea africana</i>	Cortex	<i>N. nigricollis, B. arietans</i>	71
	<i>Clutia cordata</i>	Radix	<i>N. nigricollis</i>	71
	<i>Croton draco</i>	Stems	<i>B. asper</i>	83
	<i>Croton urucurana</i>	Stem bark	<i>B. jararaca</i>	151
	<i>Emblica officinalis</i>	Roots	<i>V. russelli, E. carinatus, N. naja, Bungarus caeruleus, N. kaouthia</i>	55, 152
	<i>Euphorbia hirta</i>	Whole plant	<i>N. naja,</i>	89, 153, 154
	<i>Hevea nitida</i>	Leaf, twig	<i>B. asper</i>	74
	<i>Jatropha curcas</i>	Stem, leaf, root, latex	<i>N. naja, N. nigricollis</i>	155
	<i>Jatropha elliptica</i>	Root, stem	<i>L. muta</i>	89
	<i>Jatropha foetida</i>	Stem, leaf, root	<i>N. naja</i>	155
	<i>Jatropha gossypifolia</i>	Stem, leaf, root	<i>N. naja</i>	155
	<i>Jatropha curcas</i>	Leaf, root, stem	<i>N. naja</i>	155
	<i>Jatropha gossypifolia</i>	Leaf, root, stem	<i>B. erythromelas, B. jararaca. N. naja</i>	155- 157
	<i>Jatropha mollissima</i>	Leaf	<i>B. erythromelas, B. jararaca</i>	158
	<i>Manihot foetida</i>	Leaf, stem	<i>N. naja</i>	155
	<i>Phyllanthus emblica</i>	Root	<i>D.. russelii</i>	152, 159
	<i>Phyllanthus klotzschianus</i>	Leaves	<i>B. jaracaca</i>	114
<i>Securinega virosa</i>	Leaves	<i>N. nigricollis</i>	160	
<i>Stylosanthes erecta</i>	Folium	<i>N. nigricollis, B. arietans</i>	71	
Fabaceae	<i>Abarema cochliacarpus</i>	Stem bark	<i>B.. leucurus</i>	161
	<i>Albizia lebbeck</i>	Root/bark, seed	<i>N. kauothia, N. naja karachiensis, E. carinatus</i>	162, 163
	<i>Apuleia leiocarpa</i>	Roots	<i>B. jaracaca</i>	114
	<i>Bauhinia forficata</i>	Aerial parts	<i>B. and crotalus</i>	164
	<i>Bauhinia rufescens</i>	Seed	<i>N. nigricollis, E. ocellatus, B. arietans</i>	86
	<i>Bauhinia thonningii</i>	Cortex, radix	<i>N. nigricollis, B. arietans</i>	71
	<i>Bauhinia variegata</i>	Roots	<i>N. naja karachiensis</i>	72
	<i>Bobgunnia madagascariensis</i>	Folium, radix	<i>B. arietans, N. nigricollis</i>	71
	<i>Brongniartia</i>	Root	<i>B. atros</i>	165

	podalyrioides			
	<i>Brownea ariza</i>	Bark, leaf	<i>B. asper</i>	166
	<i>Brownea rosa-de-monte</i>	Leaf, stem bark	<i>V. russelli, B. atrox, B. asper</i>	81, 99, 102
	<i>Butea monosperma</i>	Stem bark	<i>D. russelii</i>	167
	<i>Cassia hirsuta</i>	Root	<i>N. naja</i>	168
	<i>Cassia occidentalis</i>	Leaves, roots	<i>B. moojeni</i>	169
	<i>Cassia fistula</i>	Seed	<i>B. jararaca</i>	67
	<i>Clitoria tematea</i>	Leaf	ND	69
	<i>Detarium microcarpum</i>	Leaves	<i>E. carniatus</i>	170
	<i>Dichrostachys cinerea</i>	Folium	<i>N. nigricollis, B. arietans</i>	71
	<i>Dipteryx alata</i>	Bark	<i>B. jararacussu, B. alternatus, B. asper, B. atrox,</i>	171
	<i>Entada africana</i>	Radix	<i>N. nigricollis, B. arietans</i>	71
	<i>Galactia glaucescens</i>	Leaves	<i>C. d. terrificus</i>	172
	<i>Glycyrrhiza glabra</i>	Roots	<i>B. jararaca</i>	173
	<i>Harpalyce brasiliana</i>	Roots	<i>B. jaracaca, N. naja</i>	174-176
	<i>Indigofera capitata</i>	Leaves	<i>N. nigricollis</i>	177
	<i>Indigofera pulchra</i>	Leaves	<i>N. nigricollis</i>	54
	<i>Mimosa pudica</i>	Root, herbs	<i>N. naja, N. Kaouthia, V. russelli, E. carinatus, Bungarus caeruleus, D. russelii, Malayan pit viper</i>	178-181
	<i>Mucuna pruriens</i>	Seed, leaf	<i>N. naja siamensis, N. naja, Naja hannah, Bungarus caeruleus E. carinatus,</i>	93, 182-184
	<i>Parkia bigblosa</i>	Stem bark, cortex	<i>N. nigricollis, B. arietans, E. ocellatus</i>	71, 86, 185
	<i>Pentaclethra macroloba</i>	ND	<i>B. jararacussu</i>	186
	<i>Pentaclethra macroloba</i>	Bark	<i>B. jararaca, B. jararacussu, B. moojeni, B. neuwiedi, B. pirajai</i>	186, 187
	<i>Periandra mediterranea</i>	Roots	<i>B. jaracaca</i>	114
	<i>Periandra pujalu</i>	Roots	<i>B. jaracaca</i>	114
	<i>Plathymenia reticulata</i>	Bark	<i>B. atrox, B. jararaca</i>	67, 101
	<i>Platymiscium pleiostachyum</i>	Leaves	<i>B. asper</i>	83
	<i>Psoralea corylifolia</i>	Seed	<i>N. n. karachiensis</i>	72
	<i>Psoralea corylifolia</i>	Seeds	<i>N. naja karachiensis</i>	72
	<i>Schizolobium parahyba</i>	Leaf	<i>B. alternatus, B. moojeni, B. pauloensis, C. d. terrificus</i>	188, 189
	<i>Senna auriculata</i>	Leaf	<i>E. carinatus</i>	190
	<i>Senna dariensis</i>	Whole plant	<i>B. atrox</i>	81
	<i>Stryphnodendron adstringens</i>	Root	<i>L. muta</i>	89
	<i>Tamarindus indica</i>	Root, seed, leaf,	<i>D. r. siamensis, D. russelii, N. nigricollis, B. arietans, Viper russelli</i>	71, 191-193
Gentianaceae	<i>Enicostemma hyssopifolium</i>	Whole plant	<i>N. n. karachiensis</i>	72
Heliconiaceae	<i>Heliconia curtispatha</i>	Rhizome	<i>V. russelli, B. atrox, B. asper</i>	99, 102, 103
	<i>Heliconia latispatha</i>	Rhizome	<i>B. asper</i>	194
Hymenophyllaceae	<i>Trichomanes elegans</i>	Whole plant	<i>V. russelli, B. atrox, B. asper</i>	99, 102, 103
Hypericaceae	<i>Hypericum brasiliense</i>	Whole plant	<i>B. jararaca</i>	195
	<i>Psorospermum corymbiferum</i>	Cortex, radix	<i>N. Nigricollis, B. arietans</i>	71
Hypoxidaceae	<i>Curculigo orchioides</i>	Tuber	ND	69
	<i>Milineria capitulate</i>	Folium	<i>B. arietans</i>	71
Icacinaceae	<i>Casimirella ampla</i>	Root	<i>B. atrox, B. jararaca, B. jararacussu</i>	196
	<i>Humirianthera ampla</i>	Root	<i>B. atrox, B. jararaca, B. jararacussu</i>	196
Lamiaceae	<i>Hoslundia opposita</i>	Root, leaves	<i>N. naja</i>	197
	<i>Leucas aspera</i>	Leaf, root, whole plant	<i>N. naja, D. russelii</i>	97, 198, 199
	<i>Leucas capitata</i>	Complete plant	<i>N. n. karachiensis</i>	72
	<i>Leucas cephalotes</i>	Whole plant	<i>N. n. karachiensis</i>	72
	<i>Leucas martinicensis</i>	ND	<i>N. nigricollis</i>	71
	<i>Marsypianthes ch amaedrys</i>	Inflorescence, leaf	<i>B. atrox</i>	200

	<i>Marsypianthes hptoides</i>	Herb	<i>B. jaracaca</i>	114
	<i>Ocimum tenuiflorum</i>	Whole plant	<i>N. n. karachiensis</i>	72
	<i>Peltodon radicans</i>	Flower, leaf, stem	<i>B. atrox</i>	201, 202
	<i>Rothea myricoides</i>	Cortex	<i>B. arietans, N. nigricollis</i>	71
	<i>Salvia leucantha</i>	Leaves	<i>N. n. karachiensis</i>	203
	<i>Tectona grandis</i>	Stem bark		204
	<i>Teucrium kraussii</i>	Aerial parts, cortex	<i>N. nigricollis, B. arietans</i>	71
	<i>Volkameria glabra</i>	Radix, cortex	<i>N. nigricollis, B. arietans</i>	71
Lauraceae	<i>Aniba parviflora</i>	Bark, leaf	<i>B. atrox, B. jararaca</i>	67, 101
	<i>Cassytha filiformis</i>	Herbal	<i>N. nigricollis, B. arietans</i>	71
	<i>Persea americana</i>	Seeds	<i>B. asper</i>	83
Laurencieae	<i>Laurencia aldingensis</i>	Leaves	<i>L. muta and B. jararaca</i>	205, 206
Loganiaceae	<i>Strychnos decussata</i>	Radix	<i>B. arietans</i>	71
	<i>Strychnos innocua</i>	Folium	<i>N. nigricollis, B. arietans</i>	71
	<i>Strychnos nux-vomic</i>	Seed	<i>N. kaouthia, D. russelii</i>	207
	<i>Strychnos pseudoquina</i>	Leaf	<i>B. jararaca</i>	143
	<i>Strychnos xinguensis</i>	ND	<i>B. atrox</i>	81
	<i>Struthanthus orbicularis</i>	Branch, leaf	<i>V. russelli, B. atrox, B. asper</i>	99, 102, 103
Lorasaceae	<i>Nasa speciosa</i>	Leaf	<i>B. asper</i>	113
Magnoliaceae	<i>Magnolia espinalii</i>	Leaf, twig	<i>B. asper</i>	74
	<i>Magnolia guatapensis</i>	Leaf, twig	<i>B. asper</i>	74
	<i>Magnolia hernandezii</i>	Leaf, twig	<i>B. asper</i>	74
	<i>Magnolia yarumalensis</i>	Leaf, twig	<i>B. asper</i>	74
Malpighiaceae	<i>Byrsonima crassa</i>	Leaf	<i>B. jararaca</i>	143
Malvaceae	<i>Althaea officinalis</i>	Roots	<i>Naja naja karachiensis</i>	72
	<i>Dombeya quinqueseta</i>	Cortex	<i>Naja nigricollis, B. arietans</i>	71
	<i>Grewia mollis</i>	Cortex, folium,	<i>N. nigricollis, B. arietans</i>	71
	<i>Hibiscus aethiopicus</i>	Whole plant	<i>E. carinatus, E. ocellatus, N. n. nigricollis</i>	208, 209
	<i>Pachira glabra</i>	Root bark	<i>B. pauloensis</i>	157, 210
	<i>Sterculia setigera</i>	Stem-bark , Cortex	<i>N. nigricollis, E. ocellatus and B. arietans</i>	71, 86
	<i>Waltheria indica</i>	Radix	<i>N. nigricollis, B. arietans</i>	71
Melastomataceae	<i>Bellucia dich otoma</i>	Bark	<i>B. atrox, B. jararaca</i>	67, 101, 211
	<i>Miconia albicans</i>	Stem	<i>L. muta</i>	89
	<i>Miconia fallax</i>	Stem	<i>L. muta</i>	89
	<i>Miconia sellowiana</i>	ND	<i>L. muta</i>	89
	<i>Tibouchina stenocarpa</i>	Root	<i>L. muta</i>	89
	<i>Mouriri pusa</i>	Leaf	<i>B. jararaca</i>	143
Meliaceae	<i>Azadirachta indica</i>	Leaf	<i>N. naja, V. russelli</i>	212
	<i>Carapa guianensis</i>	Leaf, twig	<i>B. asper</i>	74
	<i>Cedrela odorata</i>	Leaf, twig	<i>B. asper</i>	74
	<i>Cedrela tonduzii</i>	Leaves, stems	<i>B. asper</i>	83
	<i>Swietenia humilis</i>	Leaf, twig	<i>B. asper</i>	74
	<i>Swietenia macrophylla</i>	Leaf, twig	<i>B. asper</i>	74
	<i>Toona ciliata</i>	Root	<i>N. melanoleuca</i>	53
Menispermaceae	<i>Cissampelos mucronata</i>	Herbal	<i>N. nigricollis, B. arietans</i>	71
	<i>Cissampelos pareira</i>	Leaf	<i>B. asper</i>	213
	<i>Cyclea peltata</i>	Root	<i>N. Naja</i>	214
Moraceae	<i>Brosimum guianense</i>	Leaf	<i>B. atrox</i>	215
	<i>Castilla elastica</i>	Branch, leaf, stem	<i>B. atrox</i>	81
	<i>Dorstenia brasiliensis</i>	Roots	<i>B. jaracaca</i>	114
	<i>Ficus iteophylla</i>	Leaves	<i>N. mossandica</i>	104
	<i>Ficus nymphaeifolia</i>	ND	<i>V. russelli, B. atrox</i>	102, 99
	<i>Ficus platyphylla</i>	Folium	<i>N. nigricollis, B. arietans</i>	71
	<i>Ficus nymphaeifolia</i>	Branch, leaf, stem	<i>B. asper, B. atrox</i>	81
	<i>Morus alba</i>	Leaf, stem	<i>D. russelli, B. jaracaca</i>	216
Moringaceae	<i>Moringa oleifera</i>	Leaves	<i>N. nigricollis</i>	69, 217
Musaceae	<i>Musa paradisiaca</i>	Stem	<i>B. jararacussu, B. jararaca B. neuwiedi</i>	218, 219
Myrtaceae	<i>Myrcia guianensis</i>	Leaf	<i>B. jararaca</i>	220
	<i>Syzygium guineense</i>	Leaves	<i>B. jararaca</i>	221
Olacaceae	<i>Olax viridis</i>	Leaves	<i>B. jararaca</i>	221

Plant Extracts and Terpenes With Antivenom Properties

	<i>Ximenia americana</i>	Folium	<i>N. nigricollis, B. arietans</i>	71
Passifloraceae	<i>Passiflora quadrangularis</i>	Branch, leaf	<i>B. atrox</i>	81
Pedaliaceae	<i>Ceratotheca sesamoides</i>	Herbal	<i>N. nigricollis</i>	71
Peraceae	<i>Clusia pulchella</i>	Radix	<i>N. nigricollis</i>	71
Phyllanthaceae	<i>Flueggea virosa</i>	Radix	<i>Naja nigricollis, B. arietans</i>	71
Pinaceae	<i>Cedrus deodara</i>	Bark	<i>N. n. karachiensis</i>	72
	<i>Pinus roxburghii</i>	Oleo-resins	<i>N. naja karachiensis</i>	72
Piperaceae	<i>Piper longum</i>	Fruit	<i>D. russelii</i>	222
	<i>Piper arboreum</i>	Branch, leaf	<i>B. atrox,</i>	81
	<i>Piper pulchrum</i>	Leaf, branch, stem	<i>B. atrox</i>	81
Poaceae	<i>Coix Lacrymajobi</i>	Roots	<i>D. russelii</i>	223
	<i>Cymbopogon schoenanthus</i>	Radix	<i>N. nigricollis</i>	71
Polygalaceae	<i>Bredemeyera floribunda</i>	Roots	<i>B. jaracaca</i>	114
Polypodiaceae	<i>Pleopeltis percussa</i>	Branch, leaf, stem, whole plant	<i>B. asper, B. atrox</i>	81, 99
Primulaceae	<i>Maesa lanceolata</i>	Cortex	<i>N. nigricollis, B. arietans</i>	71
Rhamnaceae	<i>Ziziphus mucronata</i>	Radix	<i>N. nigricollis, B. arietans</i>	71
	<i>Ziziphus spina-christi</i>	Cortex	<i>N. nigricollis, B. arietans</i>	71
Rhodomelaceae	<i>Palisada flagellifera</i>	ND	<i>L. muta, B. jararaca</i>	224
Rubiaceae	<i>Crossopteryx febrifuga</i>	Cortex	<i>N. nigricollis, B. arietans</i>	71
	<i>Diodia scandens</i>	Aerial part	<i>E. carinatus</i>	225
	<i>Gonzalagunia panamensis</i>	Branch, leaf, stem	<i>B. asper, B. atrox</i>	81, 99
	<i>Ophiorrhiza mungos</i>	Root, leaf	<i>V. russelii, D. russelii</i>	226
	<i>Pentanisia prunelloides</i>	Radix	<i>N. nigricollis, B. arietans</i>	71
	<i>Pentas zanzibarica</i>	Folium	<i>N. nigricollis, B. arietans</i>	71
	<i>Randia aculeata</i>	Fruit	<i>B. asper, C. simus</i>	227
	<i>Rubia cordifolia</i>	Stem	<i>N. nigricollis, Naja naja karachiensis</i>	71, 72
	<i>Schuanniophyton magnificum</i>	Root bark	<i>N. melanoleuca, Naja kaouthia, B. gabonica</i>	228
	<i>Uncaria tomentosa</i>	Root	<i>B. asper</i>	113
	<i>Chiococca brachiata</i>	Roots	<i>B. jaracaca</i>	114
	Rutaceae	<i>Aegle marmelos</i>	Leaves, stem, root bark	<i>N. naja</i>
<i>Citrus limon</i>		Root, ripe fruits	<i>V. russelii, B. atrox, B. asper, N. karachiensis, L. muta</i>	72, 99, 102
<i>Murraya paniculata</i>		Leaf, twig	<i>B. asper</i>	74
<i>Zanthoxylum capense</i>		Radix	<i>N. nigricollis, B. arietans</i>	71
<i>Zanthoxylum monogynum</i>		Radix	<i>N. nigricollis, B. arietans</i>	230
Salicaceae	<i>Casearia sylvestris</i>	Stems, leaves, seeds, roots	<i>B. jaracaca, B. asper, B. jararacussu, B. moojeni, B. neuwiedi, B. pirajai, L. muta, N. Kaouthia</i>	114, 231, 232
	<i>Casearia grandiflora</i>	Leaf	<i>B. moojeni, B., neuwiedi</i>	233
Salvadoraceae	<i>Azima tetraacantha</i>	Leaf	<i>D. Russelii, Bungarus caeruleus</i>	2, 81, 107
Sapindaceae	<i>Billia hippocastanum</i>	Leaf, twig	<i>B. asper,</i>	74
	<i>Cupania americana</i>	Leaf, twig	<i>B. asper,</i>	74
	<i>Paullinia pinnata</i>	Root, folium	<i>N. nigricollis, B. arietans, E. carniatus</i>	71, 170
	<i>Sapindus mukorossi</i>	Fruits	<i>N. naja karachiensis</i>	72
	<i>Sapindus saponaria</i>	Whole plant,	<i>C. d. terrificus</i>	74, 234
	<i>Serjania erecta</i>	Aerial parts	<i>B. jararacussu</i>	235
Sapotaceae	<i>Manilkara subsericea</i>	Leaf, stem	<i>L. muta</i>	236
Scrophulariaceae	<i>Buddleja nitida</i>	Leaves	<i>B. asper</i>	83
Simaroubaceae	<i>Picrasma quassioides</i>	Leaves	<i>N. naja</i>	237
Siparunaceae	<i>Siparuna th ecaph ora</i>	Branch, leaf, stem	<i>B. atrox</i>	81
Solanaceae	<i>Brunfelsia unifora</i>	Leaves	<i>B. jaracaca</i>	114
	<i>Capsicum annum</i>	Ripe fruit	<i>B. atrox</i>	81
	<i>Nicotiana rustica</i>	Leaf	<i>N. nigricollis</i>	141
	<i>Schwenckia americana</i>	Folium	<i>N. nigricollis, B. arietans</i>	71
	<i>Solanum dasyphyllum</i>	Leaves, Fruits	<i>N. naja</i>	238
	<i>Solanum incanum L.</i>	Root	<i>N. naja</i>	239
	<i>Solanum xanthocarpum</i>	Roots	<i>N. naja</i>	56
	<i>Withania Somnifera</i>	Roots	<i>E. carinatus, N. naja</i>	98, 240
Symplocaceae	<i>Symplocos racemosa</i>	Bark	<i>N. naja</i>	241, 242
Theaceae	<i>Camellia sinensis</i>	Leaves	<i>N. naja kaouthia, Calloselasmahodostoma</i>	243, 244
Thymelaeaceae	<i>Gnidia anthylloides</i>	Radix	<i>N. nigricollis</i>	71
	<i>Gnidia kraussiana</i>	Radix	<i>N. nigricollis</i>	71
	<i>Gnidia splendens</i>	Radix	<i>N. nigricollis</i>	71

Urticaceae	<i>Urera baccifera</i>	Leaf	<i>B. asper</i>	113
Velloziaceae	<i>Vellozia squamata</i>	Leaf	<i>B. jararacussu</i>	245
	<i>Vernonia cinerea</i>	Whole plant		246
Verbenaceae	<i>Citharexylum macrodenium</i>	Leaves	<i>B. asper</i>	83
	<i>Clerodendrum serratum</i>	Leaves	<i>Bungarus caeruleus, D. russelli</i>	247
	<i>Clerodendrum viscosum</i>	Root	<i>N. naja</i>	248
	<i>Lantana trifolia</i>	Cortex	<i>N. nigricollis, B. arietans</i>	71
	<i>Stachytarpheta dichotoma</i>	Herb	<i>B. jaracaca</i>	114
	<i>Vitex negundo</i>	Roots, leaves	<i>V. russelli, E. carinatus, N. naja, N. kaouthia, Bungarus caeruleus, D. russelli</i>	55, 152, 249
Vitaceae	<i>Cissus populnea</i>	Stem	<i>N. nigricollis, B. arietans</i>	71
	<i>Vitis vinefera</i>	Grape eed,	<i>E. carinatus, D. russelii</i>	176, 250
Vivianiaceae	<i>Caesarea mariquitensis</i>	Leaves	<i>B. newwiedi pauloensis</i>	251
Zingiberaceae	<i>Costus Afer</i>	Leaves		252
	<i>Costus spicatus</i>	Leaf	<i>B. atrox</i>	253
	<i>Costus l sius</i>	ND	<i>B. atrox</i>	81
	<i>Curcuma longa</i>	Rhizome	<i>Naja sp, B. alternatus</i>	254, 93
	<i>Curcuma zedoaria</i>	Root	<i>D. russelii, E. carinatus, N. kaouthia, O. hannah</i>	55
	<i>Renealmia alpinia</i>	Leaf, rhizome	<i>B. asper, B. atrox</i>	81, 99, 255
	<i>urcuma aromatica</i>	Root	<i>D. russelii, E. carinatus, N. kaouthia, O. hannah</i>	55
	<i>Zingiber capitatum</i>	Rhizome	ND	69
	<i>Zingiber officinalis</i>	Rhyzome	<i>N. n. karachiensis</i>	72
Zygophyllaceae	<i>Fagonia cretica</i>	Leaf, stem	<i>N. n. karachiensis</i>	72

ND = Not described

III. Anti-Snake Venom Terpenoids From Plants

Terpenoids have been reported to have numerous pharmacological properties such as antimicrobial, antifungal, antiparasitic, antiviral, antiallergenic, antispasmodic, antihyperglycemic, anti-inflammatory, and immunomodulatory. They have also been found to be helpful in the prevention and treatment of a number of diseases, including cancer²⁵⁶. Despite the abundant ethnomedicinal and pharmacological information on the efficacy of plant extracts as antivenoms, the progress in isolation and characterization of the bioactive compounds is not impressive.

Triterpenes and sterols with antivenom properties (Table 2) include sitosterol, stigmasterol, lupeol, lupeol acetate, lupenone, 28-hydroxylupenone, betulin, betulinic acid, friedelin, β -amyrin, arjunolic acid, oleanolic acid, 11-deoxoglycyrrhetic acid, glycyrrhizin, quinovic acid, and ursolic acid^{88, 109, 230, 257}. For example, β -sitosterol and stigmasterol protected mice against lethal doses of *C. durissus terrificus*, *B. jararaca*, *B. jararacussu*, and *L. muta* venom and also reduced lethality caused by *D. russelli* and *N. kaouthia* venoms^{120, 258}. Stigmasterol exhibited inhibitory effects on enzymatic and myotoxic activities of PLA₂ from *Sapindus saponaria*²³⁴. Lupeol was found to be capable of interacting and altering the activity of the thrombin-like toxin Jararacussin-I, and capable of interacting with the BthA-I acidic PLA₂, both from *Bothrops* snakes venom^{259, 230}. Lupeol acetate inhibited lethality, hemorrhage, defibrinogenation, edema and phospholipase A₂ (PLA₂) activity induced by *Daboia russelii* venom¹⁰⁸. Lupeol acetate also inhibited lethality, cardiotoxicity and neurotoxicity in mice induced by *Naja kaouthia* venom¹⁰⁸. The triterpenoids lupeol, lupenone, 28-hydroxylupenone and betulin from *Dipteryx alata* prevented the irreversible neuromuscular blockade caused *Bothrops jararacussu* and *Crotalus durissus terrificus* venom²⁵⁹. Betulin showed the best protective effect against neuromuscular blockade and myotoxicity caused by both snake venoms²⁵⁹. Betulinic acid, oleanolic acid and ursolic acid showed the enzymatic, hemorrhagic, myotoxic and edema-inducing activities against *Bothrops atrox* venom²⁶⁰. Quinovic acid and its glycosides (quinovin glycoside C, quinovic acid-3-O- α -L-rhamnopyranoside, quinovic acid-3-O- β -D-fucopyranoside and quinovic acid-3-O- β -D-glucopyranosyl (1-4)- β -D-fucopyranoside showed significant inhibitory activity against snake venom phosphodiesterase-I^{261, 262}.

Some diterpenes have also been reported to exhibit antivenom properties including labdane lactone, labdane trialdehyde, labdane dialdehyde and corticosterone²⁶³⁻²⁶⁵. For example, labdane dialdehyde isolated from *Curcuma zedoaroides* was found active *in vitro* and *in vivo* for antivenom activity against the King cobra venom²⁶⁴.

IV. Conclusion

The results from this study show that plant possess potent snake venom neutralizing activity and could provide an alternative way to inhibit venom toxins in snakebite. A huge number of plant extracts have been

studied for antivenom properties. However, information on antivenom compounds from plants is scanty. Further research to identify the anti snake venom drugs and formulations is of importance.

Table 2: Some compound with antivenom activity against anake venoms

Compound	Plant name	Molecula Formula	Snake venom	Activity Inhibited	Ref.
β-Sitosterol	<i>Eclipta prostrata</i> , <i>Mimosa pudica</i> , <i>Pluchea indica</i> , <i>Erythroxylum subsessile</i> , <i>E. ovalifolium</i>	C ₂₉ H ₅₀ O	<i>Crotalus durissus terrificus</i> , <i>B. jararaca</i> , <i>B. jararacussu</i> , <i>L. muta</i> , <i>D. russelli</i> , <i>Deinagkistrodon acutus</i> , <i>N. kaouthia</i>	Lethality, hemorrhagic, defibrinogenation, cardiotoxicity, neurotoxicity, edematogenous, enzymatic	2, 88, 109, 120, 146, 147
Stigmasterol	<i>Cyclea peltata</i> , <i>Eclipta prostrata</i> , <i>Pluchea indica</i>	C ₂₉ H ₄₈ O	<i>C. d. terrificus</i> , <i>B. jararaca</i> , <i>B. jararacussu</i> , <i>L. muta</i> , <i>D. russelli</i> , <i>Deinagkistrodon acutus</i> , <i>N. kaouthia</i>	Lethality, hemorrhagic, defibrinogenation, cardiotoxicity, neurotoxicity, edematogenous, enzymatic	2, 88, 120, 214
Lupeol	<i>Humirianthera ampla</i> , <i>Hemidesmus indicus</i> , <i>Erythroxylum subsessile</i> , <i>E. ovalifolium</i> , <i>Zanthoxylum monogynum</i>	C ₃₀ H ₅₀ O	<i>B. atrox</i> ; <i>B. jararaca</i> ; <i>B. jararacussu</i> , <i>L. muta</i> , <i>D. russelli</i> ; <i>N. kaouthia</i>	Proteolytic, hemolytic, hemorrhagic	108, 146, 147, 196, 230, 259
Lupeol acetate	<i>H. indicus</i>	C ₃₂ H ₅₂ O ₂	<i>B. newwiedi</i> , <i>B. jararacussu</i> , <i>D. russelii</i> and <i>N. kaouthia</i>	Lethality, hemorrhagic, desfibrinogenation, edematogenous, enzymatic, cardiotoxicity, neurotoxicity	1082
Lupenone	<i>Dipteryx alata</i>	C ₃₀ H ₄₈ O	<i>B. jararacussu</i>	Myotoxicity, neurotoxicity	259
28-OH-Lupenone	<i>Dipteryx alata</i>	C ₃₀ H ₄₈ O ₂	<i>B. jararacussu</i>	Myotoxicity, neurotoxicity	259
Betulin	<i>Betula alba</i> , <i>D. alata</i>	C ₃₀ H ₅₀ O ₂	<i>B. jararacussu</i>	Myotoxicity, neurotoxicity	259, 266, 267
Betulinic acid	<i>Betula alba</i>	C ₃₀ H ₄₈ O ₃	<i>B. atrox</i>	Proteolytic enzymatic, hemorrhagic, myotoxic and edema-inducing activities	260, 266
Friedelin	<i>Albizia chevalieri</i> , <i>Erythroxylum ovalifolium</i>	C ₃₀ H ₅₀ O	<i>L. muta</i>	hemorrhagic	146, 147
β-Amyrin	<i>Apuleia leiocarpa</i>	C ₃₀ H ₅₀ O	<i>B. jararaca</i>	Lethality	109, 114
Arjunolic acid	<i>Combretum leprosum</i>	C ₃₀ H ₄₈ O ₅	<i>B. jararacussu</i> , <i>B. jararaca</i>	Lethality, hemorrhagic, myotoxicity	137
Oleanolic acid	<i>Baccharis uncinella</i>	C ₃₀ H ₄₈ O ₃	<i>D. russelii</i> , <i>N. naja</i> , <i>B. atrox</i> , <i>C. d.terrificus</i>	Enzymatic, proteolytic, hemorrhagic, edematogenous	260, 268, 269
11-deoxoglycyrrhetic acid	<i>Clematis gouriana</i>	C ₃₀ H ₄₈ O ₃	<i>D. russelii</i> , <i>N. naja</i>	-	270
Ursolic acid	<i>B. uncinella</i>	C ₃₀ H ₄₈ O ₃	<i>B. atrox</i> , <i>V. russelii</i> , <i>N. naja</i>	Proteolytic, enzymatic, edematogenous, hemorrhagic, myotoxic and edema-inducing activities	260, 268, 269
Quinovic acid	<i>Mitragyna stipulosa</i>	C ₃₀ H ₄₆ O ₅	<i>C. d.terrificus</i>	Enzymatic	261
Labdane lactone	<i>C. antinaia</i> , <i>C. contravenenum</i> , <i>C. zedoaroides</i>	C ₂₀ H ₃₀ O ₃	<i>O. hannah</i>	Lethality	263
Labdane trialdehyde	<i>C. antinaia</i> , <i>C. contravenenum</i> , <i>C. zedoaroides</i>	C ₂₀ H ₃₀ O ₃	<i>O. hannah</i>	Diaphragmatic, neurotoxic	263, 264
Labdane dialdehyde	<i>C. zedoaroides</i>	C ₂₀ H ₃₀ O ₂	<i>B. asper</i> , <i>B. atrox</i>	Lethality	264

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